Chemistry of L-Ascorbic Acid: Regioselective and Stereocontrolled 2-C- and 3-C-Allylation via Thermal Claisen Rearrangement

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We report the convenient preparation of 3-O- and 2-O-allylated derivatives of 5,6-O-isopropylidene-L-ascorbic acid (1) with various allyl substituents in high yield and their quantitative thermal Claisen rearrangement to the corresponding 2-C- and 3-C-allylated derivatives with high regio- and stereoselectivity under relatively mild conditions. This reaction will now allow the synthesis of heretofore unknown 3-C-allylated derivatives of L-ascorbic acid in high yield. The high chiral induction at the substituents of the allylic carbon, especially in the case of the 3-O-methyl-2-O-crotyl derivative, is intriguing. This general method of preparation of 2-C- and 3-C-allylated derivatives of ascorbic acid may have important applications in synthetic organic and pharmaceutical chemistry.

Introduction

Ascorbic acid, an antioxidant and radical scavenger widely distributed in aerobic organisms, is known to protect cellular components against oxidative damage by free radicals and oxidants.¹ It also serves as a reductant for several important enzymatic biotransformations.^{2,3} In addition, there is considerable evidence that biological antioxidants including ascorbic acid play an important role in the prevention of a large number of chronic diseases such as cancer, heart disease, brain dysfunction, and AIDS.⁴ Furthermore, numerous simple derivatives of ascorbic acid have been synthesized and shown to possess important pharmacological properties. For example, 5,6-O-modified ascorbic acid derivatives have been found to be clinically effective antitumor agents for various human cancers⁵ and also induce apoptosis in tumor cells.⁶ 2-Calkylated derivatives have been shown to have immunostimulant activity,7 and 2-O- and 3-O-alkylated lipid soluble derivatives are known to protect against the lipid peroxidation of the biomembrane.⁸ Therefore, the understanding of the chemistry and biochemistry of ascorbic acid and its derivatives would be helpful in the development of pharmacologically important agents with optimal activities. The chemistry of this important molecule has

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probably not been developed to expectations due to the complexity of its chemical properties.⁹

Preparation and characterization of O-alkylated and acetylated derivatives of L-ascorbic acid^{8,9} and the C-2alkylation of ascorbic acid^{10,11} with various alkylating agents have been reported. Particularly, a Michael addition of ascorbic acid with α . β -unsaturated aldehydes and ketones has been observed.^{10a-d} Moreno-Manas et al. have recently described a general method for the C-2allylation of L-ascorbic acid by allyl carbonates in the presence of palladium(0) catalyst.¹¹ However, to our knowledge there is no general method available for the synthesis of C-3-alkylated derivatives of L-ascorbic acid. In the present work, we report a general method for the preparation of both 2-C- and 3-C-allylated derivatives of L-ascorbic acid selectively in high yield via thermal Claisen rearrangements. The chemistry, stereochemistry, and spectroscopic properties of these compounds are also discussed.

Results and Discussion

Several reports have appeared in the literature recently regarding the direct C-2-alkylation of ascorbic acid with powerful alkylating agents such as allyl bromides and cinnamyl bromide in aqueous or polar solvents (e.g., DMSO), especially when the reactions were carried out for a longer period of time.¹² However, the exact mechanism for the formation of C-allylated products rather than the expected kinetically controlled, corresponding Oallylated products has not been discussed. On the other

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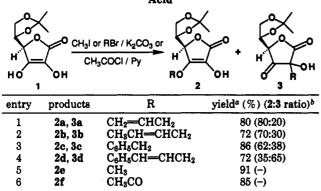
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 Table 1.
 3-O-Alkylation of 5,6-O-Isopropylideneascorbic

 Acid

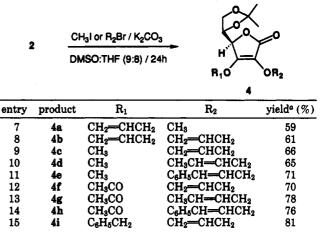


^a All the yields are given for the chromatographically purified products. ^b Calculated based on the weights of the purified material or by ¹H-NMR for nonresolvable mixtures.

hand, Kato *et al.*^{9b} have recently reported the direct O-alkylation of both 2-OH and 3-OH groups of ascorbic acid with primary alkyl halides in DMSO in the presence of K_2CO_3 or NaHCO₃, but there was no reported evidence for the formation of any C-alkylated products. Therefore, the formation of C-2-alkylated products appears to result mostly from the allylic alkylating agents.

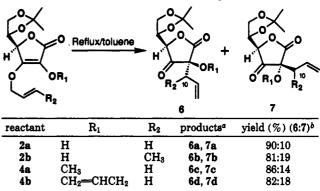
In our recent studies¹³ with several ascorbate-dependent key enzymes in the catecholamine neurotransmitter biosynthetic pathway we were interested in designing and synthesizing various ascorbate derivatives in order to probe the role of ascorbic acid in these systems. During the course of these studies it was observed that the 3-O-allyl or -crotyl derivative of 5,6-O-isopropylidene-L-ascorbic acid $(1)^{14}$ could be conveniently synthesized in high yield by reacting 1 with allyl or crotyl bromide in DMSO, in the presence of K_2CO_3 at room temperature^{8,9b} (Table 1). Although it has been previously reported^{12b} that the reaction of ascorbic acid with allyl bromide in aqueous KOH produces the 2-C-allylated product exclusively, we have observed the formation of only moderate amounts of the corresponding 2-C-allyl or -crotyl products under our reaction conditions (entries 1 and 2, Table 1). However, it was noticed that at longer reaction times and/or higher temperatures the 2-C-substituted product increased considerably and the yield of the 3-O-substituted product decreased. For example, the ¹H-NMR analysis of the crude reaction mixtures of allyl bromide and 1 showed that the yield of 2-C-allylated product was only 20% when the reaction was carried out at room temperature for 3 h and increased to 40% at 40 °C for 6 h. These results appear to indicate that the distribution of the products between 3-O- and 2-C-allylation is highly sensitive to the temperature and the reaction time. Therefore, it is possible that the kinetically favored initial 3-O-allylated product may rearrange to the thermodynamically more stable corresponding 2-C-allylated product under the reaction conditions, in a time- and temperature-dependent manner.

Heating of pure 5,6-O-isopropylidene-3-O-allyl-L-ascorbic acid (2a) in toluene to reflux for 6 h (Table 3) resulted in 100% conversion of the UV-active starting material to Table 2. 2-O-Alkylation of L-Ascorbic Acid Derivatives



 $^{\rm a}$ All the yields are given for the chromatographically purified products.

Table 3. 3-O to 2-C Rearrangement



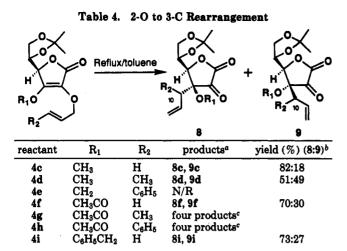
 a 100% conversion of starting material was obtained. b Calculated from distinct ¹H-NMR signals of corresponding protons.

two less polar non-UV-active products with an approximate ratio of 90:10, based on TLC and ¹H-NMR of the crude reaction mixture. The ¹H-NMR of the major product showed that the two-proton doublet at δ 4.97 of the starting material has shifted upfield to a two-proton doublet at δ 2.66, suggesting that the O-allylic methylene protons have converted to C-allylic methylene protons during the rearrangement. This was further confirmed by the upfield shift of the methylene carbon triplet from δ 72.3 to δ 39.8 in ¹³C-NMR. In addition, ¹³C-NMR of the product revealed the presence of an isolated carbonyl carbon at δ 205.5 which was absent in the starting material. These observations confirm the structure of the major product as 6a. The ¹H-NMR analysis of the partially purified minor product 7a clearly indicated that it was the C-2 diastereomer of 6a.

Pure samples of 5,6-O-isopropylidene-3-O-trans-crotyl-L-ascorbic acid (2b) and the 2-O-methylated derivative of 2a, 5,6-O-isopropylidene-2-O-methyl-3-O-allyl-L-ascorbic acid (4a), were subjected to the rearrangement under similar conditions. The ¹H-NMR analysis of the crude reaction mixture from 2b indicated that it was a mixture of only two products 6b and 7b with a ratio of 81:19 (Table 3). Both products contained an extra olefinic proton which was absent in the starting material. In addition, while the vinylic methyl group at δ 1.75 (dq) shifted upfield to δ 1.18 (d), the two-proton multiplet at δ 4.89 (O-allylic protons) collapsed to a one-proton multiplet at δ 2.70–2.80 (C-allylic proton). The presence of three olefinic protons in the ¹H-NMR and two olefinic carbons [δ 120.0(t) and δ 134.3-

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^a 100% conversion of starting material was obtained. ^b Calculated from distinct ¹H-NMR signals of corresponding protons. ^c Three products with a trace of a fourth [i.e., (10R)-8, (10S)-8, (10R)-9, and (10S)-9].

(d)] in the ¹³C-NMR of the isolated major isomer confirmed the presence of a monosubstituted terminal double bond. On the basis of this information we propose **6b** as the major product of the rearrangement of **2b**. It is also evident from the spectroscopic data that the minor product **7b** is a C-2 diastereomer of **6b**. These results together with the observation that the corresponding 3-O-benzyl derivative **2c** is resistant to the rearrangement confirm that the above reaction is a concerted Claisen rearrangement.¹⁵

We have also examined the possibility of the rearrangement of 2-O-allyl derivatives of L-ascorbic acid to yield heretofore unknown corresponding 3-C-allyl derivatives which are synthetically and pharmacologically more demanding and inaccessible by direct alkylation methods. Since the alkylation or allylation at 2-O is not practical in the presence of a more acidic 3-OH group in the molecule, we have synthesized a series of 3-O-protected 2-O-allylated ascorbic acid derivatives 4a-i (Table 2). The purified derivatives were subjected to the rearrangement under standard conditions (Table 4). In these experiments it was observed that the rearrangement is slower than that of the 3-O-allyl derivatives and was complete within 30 h giving only the expected products with no decomposition (Table 4). The rearrangement of 2,3-O-diallyl-L-ascorbate derivative 4b gave more than 97% of the 2-C-allylated products with a diastereomeric ratio of 82:18 (Table 3) again suggesting that the 3-O to 2-C rearrangement is much more favorable than the 2-O to 3-C rearrangement under competitive reaction conditions. The crotyl derivative. 5,6-O-isopropylidene-2-O-trans-crotyl-3-O-methyl-L-ascorbic acid (4d), also underwent facile rearrangement giving only two easily separable diastereomers 8d and 9d (see below). The 2-O to 3-C migration of the allyl group in these reactions is unequivocally confirmed by the presence of a carbonyl adjacent to a lactone carbonyl as indicated by the characteristic ¹³C resonance between 185 and 205 However, 5,6-O-isopropylidene-2-O-transppm.^{16a,c} cinnamyl-3-O-methyl-L-ascorbic acid (4e) was resistant to rearrangement even under vigorous conditions (refluxing in xylene for 72 h).

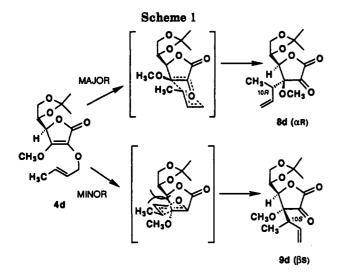
The above results clearly demonstrate that this rearrangement can be used to allylate 2-C or 3-C positions of L-ascorbic acid, selectively. In order to test whether relatively bulky 3-O blocking groups such as acetyl will retard or facilitate the reaction, 2-O-allyl (4f), 2-O-transcrotyl (4g), and 2-O-trans-cinnamyl (4h) derivatives of 5,6-O-isopropylidene-3-O-acetyl-L-ascorbic acid were prepared (Table 2), purified, and subjected to the rearrangement (Table 4). The TLC monitoring of the progress of these reactions revealed that all the reactions were much faster than the corresponding 3-O-methyl derivatives (4c, 4d, and 4e) and quantitatively produced the expected products within 4 h. The rearranged products of 4f were found to be a mixture of two diastereomers 8f and 9f with a ratio of 70:30, suggesting that even though the rearrangement is much faster, the face-selectivity is considerably less pronounced than that of the corresponding 3-Omethyl derivative. The rearrangement of 3-O-acetyl-2-O-trans-crotyl derivative 4g yielded three (with a trace of a fourth) diastereomers in a ratio of 40:40:20 based on ¹H-NMR of the crude reaction mixture (see below). In contrast to the corresponding 3-O-methyl derivative 4e, the 3-O-acetyl derivative of 2-O-trans-cinnamyl-L-ascorbic acid 4h undergoes facile rearrangement under very mild conditions (even at room temperature, 100% conversion was observed within 20 days) giving a mixture of three (with a trace of a fourth) diastereomeric products with a ratio of 50:35:15 similar to 4g. These results clearly suggest that the stereoselectivity of the rearrangement of 3-Oacetyl derivatives 4f, 4g, 4h is much less specific than that of the corresponding 3-O-methyl derivatives.

Stereochemistry. The stereochemistry of most of the rearranged products could be assigned by the close examination of their characteristic spectroscopic differences. For example, the major difference between the two products, 6a and 7a, of the rearrangement of 2a is the stereochemistry of the newly formed chiral sp³ carbon of the β -keto lactone ring. While the allyl group at C-2 and the 5,6-O-isopropylidene group containing side chain at C-4 of the ring are trans to each other in 6a, they are cis in 7a. Therefore, it could be predicted that the major influence on the ¹³C-NMR chemical shifts of the allylic methylene carbons of the two compounds should be mainly due to the differential steric interactions of these two bulky groups.^{16a} The van der Waals repulsions between the allylic methylene carbon (C-10) and the side-chain carbon (C-5) should result in the shielding of the methylene carbon of the cis product in comparison to that of the trans product.¹⁶ It is clear from the spectral data that the allylic carbon (C-10) signal of 6a is about 11 ppm more downfield than the corresponding carbon signal of 7a. Therefore, it is clear that the major product 6a of the rearrangement must have the allyl group trans to the C-4 side chain of the β -keto lactone ring which is consistent with the previous literature reports.^{10,17} Similarly, the major product 8c of the rearrangement of 4c showed a downfield shift of about 7 ppm for the C-10 signal relative to the corresponding carbon signal of the minor product 9c, confirming 8c as the major product of 2-O to 3-C rearrangement.

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The stereochemical control of the Claisen rearrangement has been extensively investigated and the stereochemical outcome of the reaction shown to be controlled by the steric factors in the transition state.^{15,18,19} It is clear that the transition states shown in Scheme 1 are the most probable for 2-O to 3-C rearrangements (similar transition states could be envisioned for 3-O to 2-C rearrangement) and the major products should be derived from the α -face alkylation. The stereochemistry at the C-10 center must also be fixed during the rearrangement (Scheme 1) since the trans-R substituent of the allyl functionality should occupy only the equatorial position in the chairlike transition state (for examples see ref 15) and should yield only two rearranged products. In agreement with these predictions prototypic compound 4d gives only two rearranged products, the absolute stereochemistry of which could be assigned as αR and βS (Scheme 1; the confirmation of these assignments is awaiting X-ray analysis). However, the behavior of 3-O-acetylated derivatives 4g and 4h is different from 4d in both the stereoselectivity (see Table 4) and the facility for the rearrangement. We believe that the behavior of these two compounds may be due to the steric and/or electronic interactions of the 3-O-acetyl group of the molecule. The details of this unexpected behavior are currently under investigation.

Experimental Section

General. Elemental analyses were performed by MHW Laboratories, Phoenix, AZ. All chemical shifts were reported on the δ (ppm) scale relative to TMS (0.00 ppm) for ¹H-NMR and to CDCl₃ (77.00 ppm) for ¹³C-NMR. Davisil grade 633 type 60A (200–425 mesh, Fisher) silica gel was used for column chromatography. TLC was performed on precoated silica gel GF plates (250 μ m, Analtech), and the compounds were observed under UV light and/or by exposure to iodine vapor. All solvents were dried with appropriate drying agents and freshly distilled. Unless otherwise specified all the products were purified by normal-phase silica gel column chromatography using a mixture of ethyl acetate and *n*-hexane with varying ratios depending on the compound.

5,6-O-Isopropylidene-L-ascorbic Acid (1). This was synthesized in 82% yield according to the procedure of Jung *et al.*:¹⁴

mp 204–206 °C (lit.⁸ mp 201–203 °C); ¹H-NMR (D₂O) δ 1.37 (6H, s), 4.17 (1H, dd, J = 5.0, 9.1 Hz), 4.31 (1H, dd, J = 7.2, 9.1 Hz), 4.59 (1H, ddd, J = 2.3, 5.0, 7.3 Hz), 4.91 (1H, d, J = 2.4 Hz); ¹³C-NMR (D₂O) δ 26.7 (q), 27.5 (q), 67.8 (t), 75.7 (d), 78.5 (d), 113.5 (s), 120.5 (s), 158.4 (s), 176.1 (s).

General Procedure for the Preparation of 2a-2e. A mixture of 1 and 1.2 equiv of K_2CO_3 in DMSO/THF (9:8) was stirred for 20 min at room temperature. The corresponding halide (1.2 equiv) in the same solvent was added dropwise, and the mixture was vigorously stirred for 4-6 h at room temperature. The reaction mixture was diluted with H_2O (4-fold) and extracted with ethyl acetate. The organic layer was thoroughly washed with H_2O , dried over anhydrous Na₂SO₄, and evaporated *in vacuo*. The products were purified by conventional silica gel column chromatography.

5,6-O-Isopropylidene-3-O-allyl-L-ascorbic Acid (2a). This was prepared from 1 and allyl bromide in 80% yield as a light greenish oil: ¹H-NMR (CDCl₃) δ 1.37 (3H, s), 1.40 (3H, s), 4.04 (1H, dd, J = 6.7, 8.6 Hz), 4.15 (1H, dd, J = 6.7, 8.6 Hz), 4.28 (1H, dt, J = 3.8, 6.6 Hz), 4.58 (1H, d, J = 3.8 Hz), 4.97 (2H, d, J = 5.7 Hz), 5.31 (1H, dq, J = 10.4, 1.3 Hz), 5.41 (1H, dq, J = 17.2, 1.4 Hz), 6.01 (1H, ddt, J = 17.2, 10.4, 5.7 Hz); ¹³C-NMR (CDCl₃) δ 25.5 (q), 25.9 (q), 65.3 (t), 72.3 (t), 74.3 (d), 75.6 (d), 110.3 (s), 119.1 (t), 119.2 (s), 132.2 (d), 148.2 (s), 171.0 (s).

5,6-O-Isopropylidene-3-*O-trans***-**crotyl**-**L**-**ascorbic Acid (2b). This was prepared from 1 and *trans***-**crotyl bromide in 72% yield as a light yellow oil: ¹H-NMR (CDCl₃) δ 1.37 (3H, s), 1.40 (3H, s), 1.75 (3H, dq, J = 6.5, 1.5 Hz), 4.02 (1H, dd, J = 6.7, 8.6 Hz), 4.13 (1H, dd, J = 6.7, 8.6 Hz), 4.26 (1H, dt, J = 3.9, 6.7 Hz), 4.55 (1H, d, J = 3.9 Hz), 4.89 (2H, m), 5.68 (1H, dtq, J = 15.3, 6.6, 1.6 Hz), 5.90 (1H, dtq, J = 15.3, 1.1, 6.5 Hz); ¹³C-NMR (CDCl₃) δ 1.7.8 (q), 25.6 (q), 25.9 (q), 65.3 (t), 72.4 (t), 74.4 (d), 75.7 (d), 110.3 (s), 119.1 (s), 125.2 (d), 132.6 (d), 148.6 (s), 171.8 (s).

5,6-O-Isopropylidene-3-O-benzyl-L-ascorbic Acid (2c). This was prepared from 1 and benzyl bromide in 86% yield as a semisolid: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.39 (3H, s), 4.02 (1H, dd, J = 6.8, 8.6 Hz), 4.10 (1H, dd, J = 6.7, 8.6 Hz), 4.26 (1H, dt, J = 3.8, 6.7 Hz), 4.57 (1H, d, J = 3.8 Hz), 5.52 (2H, two d), 7.35–7.42 (5H, m); ¹³C-NMR (CDCl₃) δ 25.5 (q), 25.9 (q), 65.3 (t), 73.5 (t), 74.2 (d), 75.7 (d), 110.3 (s), 119.5 (s), 128.4 (d), 128.6 (d), 128.7 (d), 135.7 (s), 148.6 (s), 171.1 (s).

5,6-O-Isopropylidene-3-O-methyl-L-ascorbic Acid (2e). This was prepared from 1 and methyl iodide in 91% yield as a viscous oil: ¹H-NMR (CDCl₃) δ 1.37 (3H, s), 1.40 (3H, s), 4.02 (1H, dd, J = 6.6, 8.5 Hz), 4.13 (1H, dd, J = 6.7, 8.5 Hz), 4.18 (3H, s), 4.23 (1H, dt, J = 3.8, 6.7 Hz), 4.53 (1H, d, J = 3.8 Hz).

5,6-O-Isopropylidene-3-O-acetyl-L-**ascorbic Acid (2f).** A suspension of 1 and 1.4 equiv of pyridine in CH_2Cl_2 was stirred for 20 min at room temperature, and acetyl chloride (1.2 equiv) was added dropwise. The mixture was vigorously stirred until the solution became homogeneous and was further stirred for 2 h at room temperature. The reaction mixture was diluted with H_2O (4-fold) and extracted with ethyl acetate. The organic layer was thoroughly washed with H_2O , dried over anhydrous Na₂SO₄, and evaporated *in vacuo*. The resulting residue was purified by silica gel column chromatography to yield a colorless semisolid (85%): ¹H-NMR (CDCl₃) δ 1.38 (3H, s), 1.41 (3H, s), 2.29 (3H, s), 4.41 (1H, dd, J = 6.7, 8.7 Hz), 4.20 (1H, dd, J = 6.8, 8.6 Hz), 4.44 (1H, dt, J = 3.1, 6.7 Hz), 4.71 (1H, d, J = 3.1 Hz).

General Procedure for the Preparation of 4a-4i. A solution of 2 and 1.2 equiv of K_2CO_3 in DMSO/THF (9:8) was stirred for 20 min at room temperature. The corresponding halide (1.2 equiv) in the same solvent was added dropwise, and the mixture was vigorously stirred overnight at room temperature. The reaction mixture was diluted with H_2O (4-fold) and extracted with ethyl acetate. The organic layer was thoroughly washed with H_2O , dried over anhydrous Na_2SO_4 , and evaporated *in vacuo*. The products were purified by conventional silica gel column chromatography.

5,6-O-Isopropylidene-3-O-allyl-2-O-methyl-L-ascorbic Acid (4a). This was prepared from 2a and methyl iodide in 59% yield as a semisolid: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.40 (3H, s), 3.85 (3H, s), 4.04 (1H, dd, J = 6.6, 8.5 Hz), 4.14 (1H, dd, J = 6.7, 8.5Hz), 4.30 (1H, dt, J = 3.3, 6.7 Hz), 4.53 (1H, d, J = 3.3 Hz), 4.93 (2H, dt, J = 5.6, 1.4 Hz), 5.33 (1H, dq, J = 10.5, 1.3 Hz), 5.40 (1H,

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dq, J = 17.2, 1.5 Hz), 5.98 (1H, ddt, J = 17.2, 10.5, 5.6 Hz); ¹³C-NMR (CDCl₃) δ 25.5 (q), 25.8 (q), 59.9 (q), 65.3 (t), 72.2 (t), 74.0 (d), 74.6 (d), 110.3 (s), 118.8 (t), 123.0 (s), 131.8 (d), 155.1 (s), 168.7 (s).

5.6-*O*-**Isopropylidene-2,3**-*O*-**diallyl**-L-**ascorbic** Acid (4b). This was prepared from 2a and allyl bromide in 61% yield as a light yellow oil: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.39 (3H, s), 4.04 (1H, dd, J = 6.6, 8.5 Hz), 4.14 (1H, dd, J = 6.7, 8.5 Hz), 4.30 (1H, dt, J = 3.2, 6.7 Hz), 4.55 (1H, d, J = 3.2 Hz), 4.62 (2H, two ddt), 4.94 (2H, dt, J = 5.6, 1.4 Hz), 5.27 (1H, dq, J = 10.8, 1.5 Hz), 5.39 (1H, dq, J = 17.3, 1.5 Hz), 5.38 (1H, dq, J = 17.3, 1.5 Hz), 5.98 (1H, ddt, J = 17.3, 10.5, 5.6 Hz), 5.99 (1H, ddt, J = 17.2, 10.3, 6.1 Hz); ³²C-NMR (CDCl₃) δ 25.6 (q), 25.9 (q), 65.3 (t), 72.3 (t), 72.5 (t), 74.0 (d), 74.7 (d), 110.3 (s), 118.9 (t), 119.2 (t), 121.5 (s), 131.9 (d), 132.9 (d), 155.7 (s), 168.9 (s).

5.6-O-Isopropylidene-2-O-allyl-3-O-methyl-L-ascorbic Acid (4c). This was prepared from 2e and allyl bromide in 66% yield as a light yellow viscous oil: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.39 (3H, s), 4.03 (1H, dd, J = 6.4, 8.6 Hz), 4.13 (1H, dd, J = 6.7, 8.5 Hz,), 4.15 (3H, s), 4.28 (1H, dt, J = 3.1, 6.6 Hz), 4.52 (1H, d, J = 3.1 Hz), 4.61 (2H, two ddt), 5.28 (1H, dq, J = 10.3, 1.5 Hz), 5.36 (1H, dq, J = 17.2, 1.5 Hz), 6.00 (1H, ddt, J = 17.2, 10.3, 6.2 Hz); ¹³C-NMR (CDCl₃) δ 25.5 (q), 25.8 (q), 59.6 (q), 64.2 (t), 72.7 (t), 73.8 (d), 74.5 (d), 110.3 (s), 119.3 (t), 121.5 (s), 132.8 (d), 157.1 (s), 169.9 (s).

5,6-O-Isopropylidene-2-O-trans-crotyl-3-O-methyl-L-ascorbic Acid (4d). This was prepared from **2e** and *trans*-crotyl bromide in 65% yield as a yellow viscous oil: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.39 (3H, s), 1.74 (3H, dq, J = 6.4, 1.3 Hz), 4.03 (1H, dd, J = 6.5, 8.5 Hz), 4.13 (1H, dd, J = 6.7, 8.5 Hz), 4.14 (3H, s), 4.27 (1H, dt, J = 3.1, 6.7 Hz), 4.51 (1H, d, J = 3.1 Hz), 4.52 (2H, m), 5.66 (1H, dtq, J = 15.3, 6.7, 1.5 Hz), 5.82 (1H, dtq, J = 15.3, 0.9, 6.4 Hz) ¹³C-NMR (CDCl₃): δ 17.8 (q), 25.6 (q), 25.8 (q), 59.6 (q), 65.2 (t), 72.6 (t), 73.9 (d), 74.5 (d), 110.3 (s), 121.4 (s), 125.8 (d), 132.4 (d), 157.3 (s), 169.1 (s).

5,6-O-Isopropylidene-2-O-*trans*-cinnamyl-3-O-methyl-Lascorbic Acid (4e). This was prepared from 2e and *trans*cinnamyl bromide in 71% yield as colorless crystals: mp 114-115 °C; ¹H-NMR (CDCl₃) δ 1.32 (3H, s), 1.36 (3H, s), 4.03 (1H, dd, J = 6.6, 8.5 Hz), 4.12 (1H, dd, J = 6.7, 8.5 Hz), 4.16 (3H, s), 4.27 (1H, dt, J = 3.0, 6.7 Hz), 4.51 (1H, d, J = 3.0 Hz), 4.74 (1H, ddd, J = 12.1, 6.8, 1.2 Hz), 4.81 (1H, ddd, J = 12.1, 6.6, 1.2 Hz), 6.36 (1H, dt, J = 15.9, 6.7 Hz), 6.68 (1H, d, J = 15.9 Hz), 7.23-7.36 (3H, m), 7.37-7.42 (2H, m); ¹³C-NMR (CDCl₃) δ 25.5 (q), 25.8 (q), 59.6 (q), 65.2 (t), 72.4 (t), 73.8 (d), 74.5 (d), 110.3 (s), 121.1 (s), 123.8 (d), 126.7 (d), 128.1 (d), 128.6 (d), 135.1 (d), 136.2 (s), 157.4 (s), 169.0 (s). Anal. Calcd for C₁₉H₂₂O₆: C, 65.89; H, 6.40. Found: C, 65.67; H, 6.27.

5,6-O-Isopropylidene-3-O-acetyl-2-O-allyl-L-ascorbic Acid (4f). This was prepared from 2f and allyl bromide in 70% yield as a colorless semisolid: ¹H-NMR (CDCl₈) δ 1.37 (3H, s), 1.41 (3H, s), 2.27 (3H, s), 4.08 (1H, dd, J = 6.4, 8.6 Hz), 4.16 (1H, dd, J = 6.8, 8.6 Hz), 4.38 (1H, dt, J = 3.0, 6.6 Hz), 4.69 (1H, d, J = 3.0 Hz), 4.81 (2H, two ddt), 5.35 (1H, dq, J = 10.6, 1.2 Hz), 5.40 (1H, dq, J = 18.1, 1.6 Hz), 5.95 (1H, ddt, J = 17.2, 10.5, 5.5 Hz); ¹³C-NMR (CDCl₈) δ 20.3 (q), 25.5 (q), 25.8 (q), 65.2 (t), 72.5 (t), 73.7 (d), 75.3 (d), 110.6 (s), 114.6 (s), 119.6 (t), 131.0 (d), 159.5 (s), 166.8 (s), 167.6 (s).

5,6-O-Isopropylidene-3-O-acetyl-2-O-trans-crotyl-L-ascorbic Acid (4g). This was prepared from **2f** and *trans*-crotyl bromide in 78% yield as a yellow semisolid: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.40 (3H, s), 1.76 (3H, dq, J = 6.5, 1.6 Hz), 2.27 (3H, s), 4.07 (1H, dd, J = 6.4, 8.6 Hz), 4.15 (1H, dd, J = 6.8, 8.6 Hz), 4.36 (1H, dt, J = 3.1, 6.5 Hz), 4.66 (1H, d, J = 3.1 Hz), 4.73 (2H, m), 5.62 (1H, dtq, J = 15.4, 6.4, 1.6 Hz), 5.87 (1H, dtq, J = 15.3, 1.2, 6.5 Hz); ¹³C-NMR (CDCl₃) δ 17.7 (q), 20.2 (q), 25.5 (q), 25.7 (q), 65.2 (t), 72.6 (t), 73.7 (d), 75.3 (d), 110.5 (s), 114.4 (s), 124.0 (d), 133.2 (d), 159.7 (s), 167.0 (s), 167.6 (s).

5,6-O-Isopropylidene-2-*O-trans*-cinnamyl-3-*O*-acetyl-Lascorbic Acid (4h). This was prepared from 2f and *trans*cinnamyl bromide in 76% yield as a viscous oil: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.41 (3H, s), 2.28 (3H, s), 4.09 (1H, dd, J = 6.4, 8.6Hz), 4.17 (1H, dd, J = 6.8, 8.6 Hz), 4.40 (1H, dt, J = 2.9, 6.6 Hz), 4.70 (1H, d, J = 2.9 Hz), 4.90–5.03 (2H, m), 6.29 (1H, dt, J = 15.9, 6.3 Hz), 6.71 (1H, d, J = 15.9 Hz), 7.23–7.36 (3H, m), 7.37–7.42 (2H, m); ¹³C-NMR (CDCl₃) δ 20.2 (q), 25.5 (q), 25.7 (q), 65.2 (t), 72.6 (t), 73.6 (d), 75.3 (d), 110.6 (s), 114.7 (s), 121.6 (d), 126.7 (d), 128.6 (d), 128.7 (d), 129.3 (s), 135.5 (d), 159.7 (s), 166.9 (s), 167.6 (s).

5,6-O-Isopropylidene-2-O-allyl-3-O-benzyl-L-ascorbic Acid (4i). This was prepared from 2c and allyl bromide in 81% yield as a colorless semisolid: ¹H-NMR (CDCl₃) δ 1.36 (3H, s), 1.38 (3H, s), 4.03 (1H, dd, J = 6.7, 8.6 Hz), 4.11 (1H, dd, J = 6.7, 8.5 Hz), 4.30 (1H, dt, J = 3.2, 6.7 Hz), 4.54 (2H, m), 4.55 (1H, d, J= 3.2 Hz), 5.26 (1H, dq, J = 10.3, 1.6 Hz), 5.34 (1H, dq, J = 17.2, 1.5 Hz), 5.48 (2H, br.s), 5.94 (1H, ddt, J = 17.2, 10.3, 6.1 Hz), 7.37 (5H, br.s); ¹³C-NMR (CDCl₃) δ 25.6 (q), 25.8 (q), 65.2 (t), 72.5 (t), 73.5 (d), 73.9 (t), 74.7 (d), 110.3 (s), 119.3 (t), 121.4 (s), 127.7 (d), 128.5 (d), 128.7 (d), 129.1 (s), 132.8 (d), 155.9 (s), 168.8 (s).

3-O-Acetyl-2-O-allyl-L-ascorbic Acid (5). This was obtained by acid-catalyzed removal of 5,6-O-isopropylidene group of **4f** as a colorless viscous oil: ¹H-NMR (CDCl₃) δ 2.27 (3H, s), 3.20 (2H, br s), 3.77 (1H, dd, J = 5.3, 11.5 Hz), 3.84 (1H, dd, J = 6.2, 11.5 Hz), 4.04 (1H, dt, J = 2.4, 5.7 Hz), 4.81 (2H, m), 4.86 (1H, d, J = 2.3 Hz), 5.34 (1H, dq, J = 10.5, 1.2 Hz), 5.40 (1H, dq, J = 17.3, 1.5 Hz), 5.95 (1H, ddt, J = 17.3, 10.5, 5.5 Hz); ¹³C-NMR (CDCl₃) δ 20.3 (q), 63.0 (t), 69.9 (d), 72.6 (t), 76.6 (d), 114.2 (s), 119.6 (t), 131.0 (d), 160.9 (s), 167.7 (s), 168.0 (s).

5,6-O-Isopropylidene-3-keto-2-(1-prop-2-enyl)-L-galactono- γ -lactone (6a, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged 2a: ¹H-NMR (CDCl₃) δ 1.35 (3H, s), 1.41 (3H, s), 2.66 (2H, d, J = 7.4 Hz), 4.08 (1H, dd, J = 6.9, 8.7 Hz), 4.19 (1H, dd, J = 6.9, 8.7 Hz), 4.54 (1H, dt, J = 2.0, 6.9 Hz), 4.66 (1H, d, J = 1.9 Hz), 5.26 (1H, dq, J = 16.8, 1.4 Hz), 5.28 (1H, dq, J = 10.4, 1.0 Hz), 5.69 (1H, ddt, J = 16.7, 10.4, 7.5 Hz); ¹³C-NMR (CDCl₃) δ 25.3 (q), 25.5 (q), 39.8 (t), 64.8 (t), 72.0 (s), 74.5 (d), 81.5 (d), 111.3 (s), 122.9 (t), 127.6 (d), 172.6 (s), 205.5 (s). Anal. Calcd for C₁₂H₁₆O₆: C, 56.25; H, 6.29. Found: C, 56.44; H, 6.14.

5.6-O-Isopropylidene-3-keto-2-(2-but-3-enyl)-L-galactono- γ -lactone (6b, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged 2b: ¹H-NMR (CDCl₃) δ 1.18 (3H, d, J = 6.9 Hz), 1.34 (3H, s), 1.40 (3H, s), 2.70–2.80 (1H, m), 4.07 (1H, dd, J = 6.8, 8.7 Hz), 4.18 (1H, dd, J = 6.9, 8.7 Hz), 4.53 (1H, dt, J = 2.1, 6.8 Hz), 4.60 (1H, d, J =2.2 Hz), 5.26 (1H, dt, J = 17.9, 0.9 Hz), 5.28 (1H, dt, J = 9.7, 0.9Hz), 5.75 (1H, ddd, J = 17.8, 9.7, 8.2 Hz); ¹³C-NMR (CDCl₃) δ 12.5 (q), 25.3 (q), 25.4 (q), 44.3 (d), 64.8 (t), 74.3 (s), 74.4 (d), 81.8 (d), 111.0 (s), 120.0 (t), 134.3 (d), 172.8(s), 205.7 (s).

5,6-O-Isopropylidene-3-keto-2-O-methyl-3-(1-prop-2-enyl)-L-galactono-γ-lactone (6c, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged **4a**: mp 104.5-105.5 °C; ¹H-NMR (CDCl₃) δ 1.32 (3H, s), 1.38 (3H, s), 2.63 (2H, two ddt), 3.34 (3H, s), 4.06 (1H, dd, J = 7.0, 8.5 Hz), 4.17 (1H, dd, J = 7.0, 8.5 Hz), 4.54 (1H, d, J = 1.7 Hz), 4.65 (1H, dt, J = 1.7, 6.9 Hz), 5.20 (1H, dt, J = 18.2, 1.3 Hz), 5.25 (1H, dt, J = 10.1, 0.7 Hz), 5.73 (1H, ddd, J = 18.1, 10.2, 7.9, 7.0 Hz); ¹³C-NMR (CDCl₃) δ 25.5 (q), 25.6 (q), 41.0 (t), 55.8 (q), 64.9 (t), 74.3 (d), 80.4 (s), 81.5 (d), 110.9 (s), 122.1 (t), 127.8 (d), 171.2 (s), 206.1 (s). Anal. Calcd for C₁₃H₁₈O₆: C, 57.77; H, 6.71. Found: C, 57.69; H, 6.55.

5,6-O-Isopropylidene-3-keto-2-O-(1-prop-2-enyl)-2-(1-prop-2-enyl)-L-galactono-γ-**lactone (6d, Major Isomer)**. This was isolated as a major product from the crude reaction mixture of rearranged 4b: ¹H-NMR (CDCl₃) δ 1.33 (3H, s), 1.39 (3H, s), 2.67 (2H, two ddt), 3.98 (2H, m), 4.06 (1H, dd, J = 6.9, 8.6 Hz), 4.17 (1H, dd, J = 6.9, 8.6 Hz), 4.54 (1H, d, J = 1.8 Hz), 4.64 (1H, dt, J = 1.8, 6.9 Hz), 5.19 (1H, dq, J = 10.5, 1.3 Hz), 5.20 (1H, dq, J = 16.9, 1.3 Hz), 5.25 (1H, dq, J = 10.7, 1.3 Hz), 5.30 (1H, dq, J = 17.2, 1.6 Hz), 5.74 (1H, dddd, J = 17.0, 10.1, 7.9, 6.9 Hz), 5.89 (1H, ddt, J = 17.2, 10.4, 5.7 Hz); ¹³C-NMR (CDCl₃) δ 25.4 (q), 25.7 (q), 41.2 (t), 64.9 (t), 69.2 (t), 74.2 (d), 79.9 (s), 81.5 (d), 110.9 (s), 118.0 (t), 122.2 (t), 127.8 (d), 133.2 (d), 171.4 (s), 206.1 (s). Anal. Calcd for C₁₅H₂₀O₆: C, 60.80; H, 6.80. Found: C, 60.66; H, 6.70.

5,6-O-Isopropylidene-3-keto-2-O-methyl-3-(1-prop-2-enyl)-L-**idono**- γ -**lactone (7c, Minor Isomer).** This was isolated as a minor product from the crude reaction mixture of rearranged 4a: ¹H-NMR (CDCl₃) δ 1.36 (3H, s)1.41 (3H, s), 2.73 (2H, m), 3.34 (3H, s), 4.08 (1H, dd, J = 7.1, 8.6 Hz), 4.19 (1H, dd, J = 6.8, 8.6 Hz), 4.50 (1H, dt, J = 1.9, 7.0 Hz), 4.61 (1H, d, J = 1.9 Hz), 5.15 (1H, dq, J = 17.1, 1.6 Hz), 5.21 (1H, dq, J = 10.2, 1.1 Hz), 5.88 (1H, ddt, J = 17.2, 10.1, 7.1 Hz); ¹³C-NMR (CDCl₃) δ 25.5 (q), 25.7 (q), 30.1 (t), 54.4 (q), 65.2 (t), 73.8 (d), 78.9 (s), 82.2 (d), 111.0 (s), 120.4 (t), 128.9 (d), 170.7 (s), 205.2 (s).

5.6 O Isopropylidene-2-keto-3-**O** (1-prop-2-enyl)-3-(1-prop-2-enyl)-L-galactono- γ -lactone (8b). This was isolated as a third product (less favored 2 to 3 migration product of 4b, only 2%) from the crude reaction mixture of rearranged 4b: ¹H-NMR (CDCl₃) δ 1.32 (6H, s), 2.52 (1H, dd, J = 14.8, 8.3 Hz), 2.65(1H, ddt, J = 14.9, 5.7, 1.4 Hz), 3.99 (1H, dd, J = 7.2, 8.3 Hz), 4.14 (1H, dd, J = 7.1, 8.3 Hz), 4.40 (1H, ddt, J = 12.5, 5.3, 1.5 Hz), 4.53 (1H, dt, J = 1.2, 7.1 Hz), 4.60 (1H, d, J = 1.2 Hz), 4.66 (1H, ddt, J = 12.5, 4.9, 1.7 Hz), 5.19 (1H, dq, J = 10.7, 1.2 Hz), 5.20 (1H, dq, J = 17.3, 1.7 Hz), 5.78 (1H, ddd, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1H, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1H, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 10.5, 15 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.0, 10.2, 8.4, 5.7 Hz), 5.91 (1B, ddt, J = 17.2, 10.4, 5.2 Hz); ¹³C-NMR (CDCl₃) δ 24.8 (q), 25.2 (q), 36.3 (t), 65.0 (t), 65.9 (t), 72.6 (d), 79.4 (s), 79.5 (d), 111.2 (s), 116.6 (t), 121.8 (t), 129.2 (d), 133.8 (d), 159.8 (s), 191.9 (s).

5,6-O-Isopropylidene-2-keto-3-O-methyl-3-(1-prop-2-enyl)-L-galactono- γ -lactone (8c, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged 4c: ¹H-NMR (CDCl₃) δ 1.33 (6H, s), 2.51 (1H, dd, J = 15.0, 8.3Hz), 2.66 (1H, ddt, J = 14.9, 5.6, 1.4 Hz), 3.70 (3H, s), 3.99 (1H, dd, J = 6.8, 8.4 Hz), 4.14 (1H, dd, J = 7.2, 8.4 Hz), 4.49 (1H, dt, J = 1.3, 7.0 Hz), 4.58 (1H, d, J = 1.3 Hz), 5.26 (1H, dq, J = 17.1, 1.2 Hz), 5.31 (1H, dq, J = 10.5, 1.2 Hz), 5.75 (1H, dddd, J = 17.0, 10.3, 8.3, 5.6 Hz); ¹³C-NMR (CDCl₃) δ 24.7 (q), 25.2 (q), 34.9 (t), 52.8 (q), 65.0 (t), 72.5 (d), 79.3 (s), 79.5 (d), 111.3 (s), 121.7 (t), 129.1 (d), 159.9 (s), 192.2 (s). Anal. Calcd for C₁₃H₁₈O₆: C, 57.77; H, 6.71. Found: C, 57.54; H, 6.80.

5,6-O-Isopropylidene-2-keto-3-O-methyl-3-(2-but-3-enyl)-L-galactono- γ -lactone (8d, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged 4d: mp 127-128 °C; ¹H-NMR (CDCl₃) δ 1.10 (3H, d, J = 6.8 Hz), 1.33 (6H, s), 2.70 (1H, dq, J = 9.0, 6.8 Hz), 3.73 (3H, s), 3.98 (1H, dd, J = 7.3, 8.3 Hz), 4.14 (1H, dd, J = 7.1, 8.3 Hz), 4.45 (1H, dt, J = 1.3, 7.2 Hz), 4.63 (1H, d, J = 1.3 Hz), 5.14 (1H, dt, J = 17.0, 1.3 Hz), 5.17 (1H, dt, J = 10.3, 1.3 Hz), 5.69 (1H, ddd, J = 17.0, 10.2, 9.1 Hz); ¹⁸C-NMR (CDCl₃) δ 13.6 (q), 24.9 (q), 25.4 (q), 42.3 (d), 54.6 (q), 65.2 (t), 72.9 (d), 78.5 (d), 81.1 (s), 111.2 (s), 119.3 (t), 135.9 (d), 159.8 (s), 192.6 (s). Anal. Calcd for C₁₄H₂₀O₆: C, 59.12; H, 7.09. Found: C, 58.98; H, 7.17.

5,6-O-Isopropylidene-2-keto-3-O-acetyl-3-(1-prop-2-enyl) L-galactono-γ-lactone (8f, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged **4f**: ¹H-NMR (CDCl₃) δ 1.37 (3H, s), 1.42 (3H, s), 2.16 (3H, s), 2.80 (2H, m), 4.10 (1H, dd, J = 7.3, 8.5 Hz), 4.20 (1H, dd, J = 6.9, 8.5 Hz), 4.53 (1H, dt, J = 1.6, 7.1 Hz), 4.87 (1H, d, J = 1.7 Hz), 5.19 (1H, dq, J = 17.0, 1.5 Hz), 5.26 (1H, dq, J = 10.2, 1.5 Hz), 5.90 (1H, ddt, J = 17.2, 10.1, 7.0 Hz); ¹³C-NMR (CDCl₃) δ 19.2 (q), 25.5 (q), 25.8 (q), 34.5 (t), 65.2 (t), 73.7 (d), 73.8 (s), 82.1 (d), 110.8 (s), 120.8 (t), 128.3 (d), 169.7 (s), 170.9 (s), 201.0 (s). Anal. Calcd for C₁₄H₁₈O₇: C, 56.37; H, 6.08. Found: C, 56.30; H, 6.02.

5,6-O-Isopropylidene-2-keto-3-O-acetyl-3-(2-but-2-enyl)-L-galactono- γ -lactone (8g, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged 4g: ¹H-NMR (CDCl₃) δ 1.22 (3H, d, J = 6.8 Hz), 1.38 (3H, s), 1.48 (3H, s), 2.16 (3H, s), 2.90 (1H, dq, J = 7.8, 6.8 Hz), 4.08-4.18 (2H, m), 4.46-4.55 (2H, m), 5.28 (1H, dt, J = 17.0, 1.1 Hz), 5.31 (1H, dt, J = 10.4, 1.0 Hz), 5.71 (1H, ddd, J = 17.0, 10.4, 7.8 Hz); ¹³C-NMR (CDCl₃) δ 12.3 (q), 19.2 (q), 25.3 (q), 26.7 (q), 41.4 (d), 65.2 (t), 75.1 (d), 77.3 (s), 84.8 (d), 110.0 (s), 120.6 (t), 132.9 (d), 169.3 (s), 170.3 (s), 200.9 (s).

5,6-O-İsopropylidene-2-keto-3-O-acetyl-3-(1-phenyl-1-prop-2-enyl)-L-galactono- γ -lactone (8h, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged 4h: ¹H-NMR (CDCl₃) δ 1.27 (3H, s), 1.45 (3H, s), 2.13 (3H, s), 2.97 (1H, dt, J = 6.1, 6.7 Hz), 3.82 (1H, dd, J = 6.6, 8.8 Hz), 3.97 (1H, dd, J = 7.2, 8.8 Hz), 4.05 (1H, d, J = 8.7 Hz), 4.75 (1H, d, J = 5.8 Hz), 5.17 (1H, dt, J = 17.0, 10.2, 8.6 Hz), 7.21–7.27 (2H, m), 7.32–7.40 (3H, m); ¹³C-NMR (CDCl₃) δ 19.3 (q), 25.4 (q),

26.4 (q), 51.3 (d), 64.9 (t), 72.7 (d), 76.0 (s), 83.9 (d), 110.2 (s), 119.9 (t), 128.5 (d), 129.0 (d), 129.3 (d), 129.8 (d), 132.5 (d), 134.9 (s), 169.3(s), 170.6 (s), 201.5 (s).

5,6-O-Isopropylidene-2-keto-3-(1-prop-2-enyl)-L-galactono-\gamma-lactone (8i, Major Isomer). This was isolated as a major product from the crude reaction mixture of rearranged 4i: ¹H-NMR (CDCl₃) δ 1.34 (3H, s), 1.35 (3H, s), 2.57 (1H, dd, J = 14.8, 8.5 Hz), 2.75 (1H, ddt, J = 14.8, 5.5, 1.4 Hz), 4.00 (1H, dd, J = 7.3, 8.3 Hz), 4.11 (1H, dd, J = 7.1, 8.3 Hz), 4.55 (1H, dt, J = 1.2, 7.2 Hz), 4.63 (1H, d, J = 1.2 Hz), 4.90 (1H, d, J = 10.9 Hz), 5.25 (1H, dm, J = 10.5 Hz), 5.30 (1H, d, J = 10.8 Hz), 5.31 (1H, d, J = 18.6 Hz), 5.82 (1H, dddd, J = 18.8, 10.3, 8.5, 5.5 Hz), 7.30 (5H, br s); ¹³C-NMR (CDCl₃) δ 24.9 (q), 25.4 (q), 36.3 (t), 64.9 (t), 67.1 (t), 72.7 (d), 79.3 (s), 79.4 (d), 111.2 (s), 121.9 (t), 127.5 (d), 127.9 (d), 128.4 (d), 129.2 (s), 137.5 (d), 159.8 (s), 192.2 (s).

5.6-O-Isopropylidene-2-keto-3-O-methyl-3-(1-prop-2-enyl)-L-idono- γ -lactone (9c, Minor Isomer). This was isolated as a minor product from the crude reaction mixture of rearranged 4c: ¹H-NMR (CDCl₃) δ 1.29 (3H, s), 1.31 (3H, s), 2.46 (1H, ddt, J =17.0, 8.2, 1.1 Hz), 3.00 (1H, ddt, J = 17.0, 5.2,2.0 Hz), 3.19 (3H, s), 4.04 (1H, dd, J = 6.9, 8.4 Hz), 4.12 (1H, dd, J = 6.9, 8.4 Hz), 4.41 (1H, dt, J = 0.9, 6.9 Hz), 4.61 (1H, d, J = 0.9 Hz), 5.30 (1H, dq, J = 10.3, 1.0 Hz), 5.33 (1H, dq, J = 17.3, 2.0 Hz), 5.33 (1H, dddd, J = 17.3 10.4, 8.2, 5.2 Hz); ¹³C-NMR (CDCl₃) δ 24.8 (q), 25.2 (q), 27.6 (t), 51.2 (q), 64.9 (t), 72.7 (d), 77.2 (s), 82.6 (d), 111.4 (s), 120.3 (t), 129.5 (d), 160.0 (s), 186.0 (s).

5.6-O-Isopropylidene-2-keto-3-O-methyl-3-(2-but-3-enyl)-L-idono- γ -lactone (9d, Minor Isomer). This was isolated as a minor product from the crude reaction mixture of rearranged 4d: ¹H-NMR (CDCl₃) δ 1.11 (3H, d, J = 6.9 Hz), 1.33 (6H, s), 2.89 (1H, pentet, J = 1.2, 6.9 Hz), 3.70 (3H, s), 3.98 (1H, dd, J = 7.2, 8.3 Hz), 4.14 (1H, dd, J = 7.1, 8.3 Hz), 4.47 (1H, dt, J = 1.4, 7.1 Hz), 4.58 (1H, d, J = 1.4 Hz), 5.17 (1H, dt, J = 17.3, 1.3 Hz), 5.24 (1H, dt, J = 10.5, 1.1 Hz), 5.81 (1H, ddd, J = 17.3, 10.5, 6.9 Hz); ¹⁸C-NMR (CDCl₃) δ 13.2 (q), 24.9 (q), 25.4 (q), 41.7 (d), 53.9 (q), 65.2 (t), 73.0 (d), 79.2 (d), 80.9 (s), 111.1 (s), 118.3 (t), 135.6 (d), 159.8 (s), 193.1 (s).

5,6-O-Isopropylidene-2-keto-3-O-acetyl-3-(1-prop-2-enyl) L-idono-\gamma-lactone (9f, Minor Isomer). This was isolated as a minor product from the crude reaction mixture of rearranged **4f**: ¹H-NMR (CDCl₃) δ 1.38 (3H, s), 1.48 (3H, s), 2.16 (3H, s), 2.68 (1H, dd, J = 13.6, 8.0 Hz), 2.76 (1H, ddq, J = 13.6, 6.9, 1.1 Hz), 4.10 (1H, dd, J = 7.3, 8.5 Hz), 4.20 (1H, dd, J = 6.9, 8.5 Hz), 4.53 (1H, dt, J = 1.6, 7.1 Hz), 4.56 (1H, d, J = 1.7 Hz), 5.29 (1H, dq, J = 17.0, 1.3 Hz), 5.33 (1H, dq, J = 10.4, 1.1 Hz), 5.72 (1H, dddd, J = 17.0, 10.2, 7.9, 6.9 Hz); ¹³C-NMR (CDCl₃) δ 19.2 (q), 25.3 (q), 26.7 (q), 37.1 (t), 65.1 (t), 73.8 (s), 75.1 (d), 84.7 (d), 110.1 (s), 123.3 (t), 126.4 (d), 169.2(s), 170.3 (s), 201.3 (s).

5,6-O-Isopropylidene-2-keto-3-O-acetyl-3-(1-phenyl-1-prop-2-enyl)-L-galactono- γ -lactone (9h, Second Major Isomer). This was isolated as the second major product from the crude reaction mixture of rearranged 4h: ¹H-NMR (CDCl₃) δ 1.29 (6H, s), 2.15 (3H, s), 3.15 (1H, d, J = 8.3 Hz), 3.88 (1H, d, J = 8.4 Hz), 3.91 (1H, dd, J = 5.9, 9.2 Hz), 3.99 (1H, dd, J = 6.4, 9.2 Hz), 4.37 (1H, dt, J = 8.3, 6.2 Hz), 5.28 (1H, dt, J = 16.9, 1.2 Hz), 5.36 (1H, dt, J = 10.3, 1.0 Hz), 6.42 (1H, ddd, J = 17.0, 10.3, 8.4 Hz), 7.17-7.22 (2H, m), 7.32-7.40 (3H, m); ¹³C-NMR (CDCl₃) δ 19.2 (q), 25.2 (q), 26.5 (q), 53.5 (d), 65.0 (t), 74.7 (d), 77.1 (s), 84.6 (d), 109.7 (s), 120.5 (t), 128.7 (d), 129.0 (d), 129.3 (d), 132.0 (d), 133.4 (s), 168.3(s), 170.3 (s), 202.1 (s).

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Supplementary Material Available: Copies of ¹H and ¹³C NMR spectra of many compounds (36 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.